# TRITERPENES FROM DOUGLAS FIR SAPWOOD

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Abstract—The saponified ether-soluble extractives of Douglas fir sapwood contained (24R)- $4\alpha$ ,  $14\alpha$ , 24-trimethyl- $9\beta$ , 19-cyclo- $5\alpha$ -cholestan- $3\beta$ -ol (24R-cycloeucalanol), a new natural product;  $4\alpha$ ,  $14\alpha$ -dimethyl- $9\beta$ , 19-cyclo-24-methylene- $5\alpha$ -cholestan- $3\beta$ -ol (cycloeucalenol); and (24R)- $4\alpha$ , 24-dimethyl- $5\alpha$ -cholest-7-en- $3\beta$ -ol (24R-methyllophenol); this is the first time they have been reported from Douglas fir.

### INTRODUCTION

Douglas fir [Pseudotsuga menziesii (Mirb.) Franco] is a major conifer species used by many kraft mills in western United States and Canada for producing pulp. Tall oil, a commercially important chemical by-product of kraft pulping, originates from extractable components of wood. While investigating Douglas fir extractives and their relation to tall oil obtainable from wood [1], an unidentified triterpene alcohol (ca 3-4% of the Et<sub>2</sub>O extract) was isolated from the non-saponifiable fraction. Further investigation has shown this material is a mixture of three triterpenes: 24R-cycloeucalanol [(24R)- $4\alpha$ ,  $14\alpha$ , 24-trimethyl- $9\beta$ , 19-cyclo- $5\alpha$ -cholestan- $3\beta$ -ol (1a), the major constituent, and cycloeucalenol  $[4\alpha, 14\alpha]$ dimethyl-9 $\beta$ ,19-cyclo-24-methylene-5 $\alpha$ -cholestan-3 $\beta$ -ol (2a) and 24R-methyllophenol [(24R)-4 $\alpha$ ,24-dimethyl-5 $\alpha$ cholest-7-en-3 $\beta$ -ol] (3a), both minor constituents. This is the first reported occurrence of cycloeucalanol as a natural product, and of cycloeucalenol and 24methyllophenol from Douglas fir.

### RESULTS AND DISCUSSION

24R-Cycloeucalanol (1a), cycloeucalenol (2a) and 24Rmethyllophenol (3a) were isolated and characterized as their respective acetates (1b, 2b and 3b). 24R,S-Cycloeucalanyl acetate has been previously synthesized by hydrogenation of cycloeucalenyl acetate [2]. This is the first reported occurrence of 24R-cycloeucalanyl acetate as a pure diastereoisomer and of 24Rcycloeucalanol as a natural product. Cycloeucalenol has been isolated from numerous natural sources, and is a postulated intermediate in the biosynthesis of phytosterols from squalene [3]. 24ξ-Methyllophenol has been isolated from Larix decidua leaves [4], from Sorghum vulgare grain [5, 6], Digitalis pupurea seeds [7], avocado oil [8] and grapefruit peel [9]; however, in each isolation the proof of structure was based solely on GC/MS data. This is the first report of cycloeucalenol and 24R-methyllophenol as natural products from Douglas

## 24R-Cycloeucalanyl acetate (1b)

The molecular formula of this compound is  $C_{32}H_{54}O_2$  as determined by high-resolution MS. The MS

fragmentation pattern (Fig. 1) is consistent with the proposed structure and with MS data reported for compounds of this type [10]. The ions at m/z 343 [M<sup>+</sup> – side chain (C<sub>9</sub>H<sub>19</sub>)], m/z 302 and m/z 175 [m/z 302 – side chain (C<sub>9</sub>H<sub>19</sub>)] support the presence of the C(24)-Me. The <sup>1</sup>H NMR (Table 1) is also consistent with the proposed structure (**1b**). From the position of the 9 $\beta$ ,19-cyclopropane AB Hs, the methyl at C(4) must have the 4 $\alpha$ -configuration [11]. The C(24)-Me is confirmed by the magnetic nonequivalence of the isopropyl methyls [C(26) and C(27)] in the <sup>1</sup>H NMR because of the asymmetric center at C(24) [12]. The chemical shifts for the C(26), C(27) and C(28) protons are the same as those reported for similar compounds having the 24R-configuration [13].

Optical rotation measurements also establish the stereochemistry of cycloeucalanyl acetate as 24R. The molecular rotation ( $[M]_D$ ) of a 24R-Me compound is ca  $29^\circ$  more dextrorotatory than is the corresponding 24-H compound; the 24S-Me compound is about  $29^\circ$  more levorotatory [14]. The  $[M]_D$  of cycloeucalanyl acetate (16) is  $+333^\circ$ ; the  $[M]_D$  of 31-norcycloartanyl acetate (16), the corresponding 16-H compound, is 16-Nus, the 16-D of cycloeucalanyl acetate is about 16-Minor dextrorotatory than is the 16-norcycloartanyl acetate, as expected for the 16-Me configuration.

The identity of this compound was further confirmed by comparison with 24R, S-cycloeucalanyl acetate (4b) obtained by hydrogenation of cycloeucalenyl acetate (2b). The two materials were identical by TLC and GLC. In addition, the  $^1H$  NMR spectra were identical; however, the relative intensities of various peaks in the methyl region of the spectra (ca  $\delta$  0.75–1.00) were not the same. This difference is expected because 24R, S-cycloeucalanyl acetate is a mixture of C(24)-epimers in which the chemical shifts of the C(26), C(27), C(28) and possibly the C(21) protons, are slightly different because of the asymmetric center at C(24).

## Cycloeucalenyl acetate (2b)

The compound isolated from Douglas fir was identical to authentic cycloeucalenyl acetate by mp,  $[\alpha]_D$ , NMR, IR, TLC, GLC and MS.

Fig. 1. Mass spectral El fragmentation of 24R-cycloeucalanyl acetate.

## 24R-Methyllophenyl acetate (3b)

The high-resolution MS indicated a molecular formula of  $C_{31}H_{52}O_2$ . The MS (Fig. 2) contained peaks with the approximate relative intensity as those reported for  $24\xi$ -methyllophenyl acetate [5]. The peaks at m/z 329, 269, and 161 indicate a C(24)-Me in the side chain. The low intensity of the m/z 288 peak, formed by retro-Diels-Alder from  $M^+$ , is expected for the  $5\alpha$ - $\Delta$ <sup>7</sup> configuration [16].

The <sup>1</sup>H NMR data are also consistent with the proposed structure (Table 1). The chemical shifts of the =CH at  $\delta$  5.21 and of the C(18) protons at  $\delta$  0.53 are fully consistent with a  $\Delta$ <sup>7</sup> double bond. The chemical shifts for the C(26), C(27), and C(28) protons are the same as those reported for similar compounds having the 24R configuration [13].

Biogenetically, the stereochemistry at C(24) should be the same as that for 24R-cycloeucalanyl acetate. Because this compound was obtained only in about 90% purity, the optical rotation must be interpreted with caution to establish the stereochemistry at C(24). The  $[M]_D$  is about 52° more dextrorotatory than that of lophenyl acetate (5) [17]; this difference is comparable to the 41° difference between cholest-7-en-3 $\beta$ -yl acetate ( $[M]_D + 18^\circ$ ) and ergost-7-en-3 $\beta$ -yl acetate (24S-Me;  $[M]_D - 23^\circ$ ) [18], but is larger than the 29° expected. However, the direction of the  $\Delta[M]_D$  is consistent with the 24R configuration.

The identity of this compound was further confirmed by comparing its hydrogenation product [24R-methyllophanyl acetate (7)] with 24R,S-methyllophanyl acetate (6) obtained by hydrogenation of 24-methylenelophenyl acetate (5). The 24R- and 24R,S-methyllophanyl acetates were identical by TLC and GLC. In addition, the <sup>1</sup>H NMR spectra were identical; however, the relative intensities of various peaks in the methyl region between ca  $\delta$ 0.75–1.00 were not the same. This difference is expected because 24R,S-methyllophanyl acetate is a mixture of the C(24)-epimers.

### **EXPERIMENTAL**

Mps were measured in evacuated capillaries and are corrected.  $^1H$  NMR spectra were determined in CHCl<sub>3</sub>. GLCs (Table 2) were obtained with a 10-m glass capillary coated with SP 2100 operated at 250°. TLCs were obtained on Si gel impregnated with 20 % AgNO<sub>3</sub> using petrol (PE)–Et<sub>2</sub>O (90:10).

Initial isolation of triterpenes. Chips of Douglas fir sapwood (ca 1.5 kg) were ground in a Wiley Mill, and the resulting milled wood continuously extracted with Et<sub>2</sub>O in a Soxhlet apparatus for 1 week. The Et<sub>2</sub>O soln was concd, extracted with dil. NaOH to remove the acidic components, evapd to dryness to yield the neutrals, and saponified by refluxing with KOH in EtOH for 4 hr. The EtOH was evapd and the residue taken up in Et<sub>2</sub>O and extracted with dilute NaOH to remove any acidic components. The Et<sub>2</sub>O was evapd to yield 1.3 g of non-saponifiable neutrals.

The non-saponifiables were chromatographed over Si gel with toluene and toluene–Et<sub>2</sub>O mixtures. Toluene–Et<sub>2</sub>O (95:5 and 90:10) eluted a mixture of 24-methylenecycloartanol and cycloartenol identified by TLC, GLC and <sup>1</sup>H NMR (60 MHz). Further elution gave a mixture of cycloeucalanol, cycloeucalenol and 24-methyllophenol, followed by a mixture of campesterol, stigmastanol and sitosterol. The center portion of the fractions containing cycloeucalanol, cycloeucalenol and 24-methyllophenol (127 mg) was acetylated with Ac<sub>2</sub>O-pyridine. The mixed acetates were chromatographed over Si gel–20 % AgNO<sub>3</sub>. PE and PE–Et<sub>2</sub>O (99:1 and 99:2) eluted 79 mg of a mixture of cycloeucalanyl acetate and 24-methyllophenyl acetate. PE–Et<sub>2</sub>O (95:5) eluted 35 mg of impure cycloeucalenyl acetate.

0.92, 3 H, d, J = 7 Hz

0.84, 3 H, d, J = 7 Hz

0.80, 3 H, d, J = 7 Hz

0.84, 3 H, d, J = 7 Hz

0.77, 3 H, d, J = 7 Hz

0.71, 3 H, s

0.82, 3 H, s

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Proton	24R-Cycloeucalanyl acetate (1b)	Cycloeucalenyl acetate (2b)	24R-Methyllophenyl acetate(3b)	24R-Methyllophanyl acetate (7)
[	4.50, 1 H, m	4.50, 1 H, m	4.43, 1 H, m	4.46, 1 H, m
O     -C-CH <sub>3</sub>	2.04, 3 H, s	2.04, 3 H, s	2.04, 3 H, s	2.04, 3 H, s

0.89, 3 H, d, J = 7 Hz

0.90, 3 H, s

0.96, 3 H, s

 $J = 4 \,\mathrm{Hz}$ 

4.66, 1 H, s 4.71, 1 H, s

0.27, 2 H, ABdd

 $(\delta_{\rm A} = 0.14, \, \delta_{\rm B} = 0.39)$ 

0.83, 3 H, d, J = 7 Hz

1.015, 3 H, d, J = 7 Hz

1.020, 3 H, d, J = 7 Hz

Table 1. <sup>1</sup>H NMR spectral data of Douglas fir triterpenes\*

0.88, 3 H, d, J = 8 Hz

0.90, 3 H, s

0.96, 3 H, s

 $J = 4 \,\mathrm{Hz}$ 

0.27, 2 H, ABdd

 $(\delta_{A} = 0.15, \delta_{B} = 0.40)$ 

 $0.85, 3 \, H, d, J = 7 \, Hz$ 

 $0.80, 3 \,\mathrm{H}, d, J = 7 \,\mathrm{Hz}$ 

0.84, 3 H, d, J = 7 Hz

0.78, 3 H, d, J = 7 Hz

C(3) - H

C(3) - O - C

 $C(4\alpha)-CH_3$ 

 $C(14) - CH_3$ 

 $C(18) - H_3$ 

C(19)-H

 $C(21)-H_3$ 

 $C(26)-H_3$ 

 $C(27)-H_3$ 

 $C(28) - H_3$ 

C(7)-H

Fig. 2. Mass spectral EI fragmentation of 24R-methyllophenyl

24R-Cycloeucalanyl acetate (1b). The mixture of cycloeucalanyl acetate and 24-methyllophenyl acetate was rechromatographed over 40 g Si gel-20 % AgNO<sub>3</sub>. PE-Et<sub>2</sub>O (98:2) eluted 51 mg of cycloeucalanyl acetate that was crystallized from MeOH: mp  $107.5-108^{\circ}$ ,  $[\alpha]_{D}^{25} + 70.9^{\circ}$  (c 0.9). Reported for 24R, S-cycloeucalanyl acetate [2]: mp 112–113°, [ $\alpha$ ]<sub>D</sub> +62°. This material was homogeneous by TLC and GLC.  $v_{max}^{KBr}$  cm<sup>-1</sup>: 1735 and 1252 (acetate). NMR—see Table 1. MS (probe, 175°, 60 eV) m/z (rel. int.): 470 (10; M<sup>+</sup>; C<sub>32</sub>H<sub>54</sub>O<sub>2</sub>), 455 (10; C<sub>31</sub>H<sub>52</sub>O<sub>2</sub>), 411

(31), 410 (100;  $C_{30}H_{50}$ ), 396 (22;  $C_{29}H_{48}$ ), 395 (76;  $C_{29}H_{47}$ ), 380.9 (M\*, 410  $\rightarrow$  395), 358 (M\*; 470  $\rightarrow$  410), 355 (14;  $C_{26}H_{43}$ ), 343 (M\*; 455  $\rightarrow$  395), 302 (18;  $C_{22}H_{38}$ ), 283 (28;  $C_{21}H_{31}$ ), 233.5  $(M^*; 343 \rightarrow 283), 220 (13; C_{16}H_{28}), 189 (18; C_{14}H_{21}), 188 (15;$  $C_{14}H_{20}$ ), 175 (29;  $C_{13}H_{19}$ ), 173 (16;  $C_{13}H_{17}$ ), 163 (22;  $C_{12}H_{19}$ ).  $161 (24; C_{12}H_{17}), 149 (15; C_{11}H_{17}), 148 (11; C_{11}H_{16}), 147 (24;$  $C_{11}H_{15}$ ), 145 (14;  $C_{11}H_{13}$ ), 137 (11;  $C_{10}H_{17}$ ), 136 (16;  $C_{10}H_{16}$ ), 135 (28;  $C_{10}H_{15}$ ), 134 (16;  $C_{10}H_{14}$ ), 133 (25;  $C_{10}H_{13}$ ), 131 (12;  $C_{10}H_{11}$ ), 123 (19;  $C_{9}H_{5}$ ), 121 (35;  $C_{9}H_{13}$ ), 120 (21;  $C_{9}H_{12}$ ), 119  $(29; C_9H_{11}), 109 (39; C_8H_{13}), 108 (21; C_8H_{12}), 107 (41; C_8H_{11}),$ 105 (26).  $M^+$  m/z 470.4099. Required for  $C_{32}H_{54}O_2$ ,  $M^+$  m/z 470.4122.

0.92, 3 H, d, J = 7 Hz

0.85, 3 H, d, J = 7 Hz

0.80, 3 H, d, J = 7 Hz

0.84, 3 H, d, J = 7 Hz

0.78, 3 H, d, J = 7 Hz

5.21, 1 H, br. d,  $J = 5 \,\mathrm{Hz}$ 

0.53, 3 H, s

0.83, 3 H, s

24R-Methyllophenyl acetate (3b). Continued elution of the previous column with PE-Et<sub>2</sub>O (98:2) gave 7 mg of a compound that was 80% pure by GLC. This material was cryst. from methanol to give 3 mg of 24R-methyllophenyl acetate (+90%)purity): mp 135–135.5°,  $[\alpha]_D^{25} + 38.6^\circ$  (c 0.3). NMR—see Table 1. MS (probe, 220°, 70 eV) m/z (rel. int.): 456 (100; M<sup>+</sup>, C<sub>31</sub>H<sub>52</sub>O<sub>2</sub>), 441 (20;  $M^+ = Me$ ,  $C_{30}H_{49}O_2$ ); 396 (15;  $M^- = HOAc$ ,  $C_{29}H_{49}$ ), 381 (19;  $M^+ - Me - HOAc$ ,  $C_{28}H_{45}$ ), 329 (8;  $M^+ - C_9H_{19}$ ), 288 (4;  $C_{21}H_{36}$ ), 287 (9;  $C_{18}H_{39}O_2$ ), 270 (14;  $C_{20}H_{30}$ ), 269 (67,  $M^+ - HOAc - C_9H_{19}$ ), 243 (23;  $C_{18}H_{27}$ ), 227 (41;  $C_{17}H_{23}$ ), 161 (20;  $C_{12}H_{17}$ ). M + m/z 456.3943. Required for  $C_{31}H_{52}O_2$  M + m/z 456.3966. Reported MS [5] m/z (rel. int.): 456 (100), 381 (25), 329 (7), 287 (9), 269 (90), 227 (45).

Cycloeucalenyl acetate (2b). The impure sample of cycloeucalenyl acetate was chromatographed over Si gel-20% AgNO<sub>3</sub>. PE-Et<sub>2</sub>O (90:10) eluted 20 mg of cycloeucalenyl acetate that was cryst. from methanol: mp  $108-109^{\circ}$ ,  $[\alpha]_D^{25} + 62.2^{\circ}$ (c 0.9). Reported: mp 110°,  $[\alpha]_D + 63^\circ$  [2]; mp 105–109°,  $[\alpha]_D$ +61.6° [19]. NMR—see Table 1. This compound was identical to authentic cycloeucalenyl acetate by NMR, IR, TLC and GLC. MS (probe,  $200^{\circ}$ , 70 eV) m/z (rel. int.): 468 (8; M<sup>+</sup>), 453 (9; M<sup>+</sup> - Me), 425 (4), 409 (36), 408 (100; M<sup>+</sup> - HOAc), 394 (27), 393 (85), 365 (7), 353 (12), 325 (8), 324 (5), 300 (24), 283 (22), 281 (15). Reported MS m/z (rel. int.) [19]: 468 (12), 453 (9), 425 (5), 408 (100), 393 (52), 365 (5), 353 (9), 325 (7), 324 (5), 300 (12), 283 (14), 281 (10).

<sup>\*</sup>At 270 MHz; results given in  $\delta$  (ppm).

Table 2. GLC of Douglas fir triterpenes

Compound	RRT*
24R-Methyllophanyl acetate (7)	0.92
Sitosteryl acetate	1.00
24R-Methyllophenyl acetate (3b)	1.03
Cycloeucalenyl acetate (2b)	1.05
24R-Cycloeucalanyl acetate (1b)	1.07

<sup>\*</sup>RRT, relative retention time compared to sitosteryl acetate.

24R,S-Cycloeucalanyl acetate (4b) from cycloeucalenyl acetate (2b). Cycloeucalenyl acetate (14 mg) in dimethoxyethane (20 ml) was hydrogenated over 10 % Pd–C (20 mg) for 2 hr at room temp. The reaction mixture was filtered through a small bed of Si gel with Et<sub>2</sub>O washings. The solvent was evapd in vacuo to yield 24R,S-cycloeucalanyl acetate (13 mg). This material was chromatographed over Si gel. PE–Et<sub>2</sub>O (95:5) eluted chromatographically pure material identical with 24R-cycloeucalanyl acetate by TLC and GLC. This compound was cryst. from MeOH: mp 103–104°,  $[\alpha]_D^{24}$  +66° (c 0.5). Reported [2]: mp 112–113°,  $[\alpha]_D$  +62°.

24R-Methyllophanyl acetate (7) from 24R-methyllophanyl acetate (3b). 24R-Methyllophenyl acetate (2 mg) in dimethoxyethane (20 ml) was hydrogenated over 10% Pd-C (15 mg) for 3 hr at room temp. The reaction mixture was filtered through a small bed of Si gel with Et<sub>2</sub>O washings. The Et<sub>2</sub>O was evapd to give 24R-methyllophanyl acetate (7) (2 mg) that was chromatographed over Si gel. PE-Et<sub>2</sub>O (98:2) eluted 2 mg of material:  $\lceil \alpha \rceil_D^{2.3} + 46^\circ$  (c 0.1). NMR—see Table 1.

24R,S-Methyllophanyl acetate (6) from 24-methylenelophenyl acetate (5). 24-Methylenelophenyl acetate [20] (14 mg) in dimethoxyethane (20 ml) was hydrogenated over 10% Pd-C (10 mg) at room temp. for 1.5 hr. The reaction mixture was filtered through a small bed of Si gel with Et<sub>2</sub>O washings.

The Et<sub>2</sub>O was evapd to give 24R,S-methyllophanyl acetate (14 mg). This material was chromatographed over Si gel. PE-Et<sub>2</sub>O (98:2) eluted 8 mg of a compound that was cryst. from MeOH: mp 106-107°,  $[\alpha]_0^{24} + 36^{\circ}$  (c 0.4). This compound was identical to 24R-methyllophanyl acetate by TLC and GLC.

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### REFERENCES

- Foster, D. O., Zinkel, D. F. and Conner, A. H. (1980) Tappi 63, 103.
- Cox, J. S. G., King, F. E. and King, T. J. (1956) J. Chem. Soc. 1384.
- Goad, L. J. and Goodwin, T. W. (1972) in *Progress in Phytochemistry* (Reinhold, L. and Liwschitz, Y., eds.) Vol. 3, p. 113. Interscience, New York.
- 4. Goad, L. J. and Goodwin, T. W. (1967) Eur. J. Biochem. 1, 357
- Bowden, B. N. and Palmer, M. A. (1975) Phytochemistry 14, 1140
- Palmer, M. A. and Bowden, B. N. (1975) Phytochemistry 14, 2049.
- 7. Evans, F. J. (1972) J. Pharm. Pharmacol. 24, 227.
- Itoh, T., Tamura, T., Matsumoto, T. and Dupaigne, P. (1976) Fruits 31, 473.
- Williams, B. L., Goad, L. J. and Goodwin, T. W. (1967) *Phytochemistry* 6, 1137.
- 10. Aplin, R. T. and Hornby, G. M. (1966) *J. Chem. Soc. B*, 1078.
- Berti, G., Bottari, F., Macchia, B., Marsili, A., Ourisson, G. and Piotrowska, H. (1964) Bull. Soc. Chim. Fr. 2361.
- Whitesides, G. M., Holtz, D. and Roberts, J. D. (1964) J. Am. Chem. Soc. 86, 2628.
- Rubinstein, I., Goad, L. J., Clague, A. D. H. and Mulheirn, L. (1976) Phytochemistry 15, 195.
- 14. Bergmann, E. and Low, E. M. (1947) J. Org. Chem. 12, 67.
- Devys, M., Andre, D. and Barbier, M. (1969) C.R. Acad. Sci. Sect. D 269, 798.
- Dixon, J. S., Midgley, I. and Djerassi, C. (1977) J. Am. Chem. Soc. 99, 3432.
- 17. Ogunkoya, L. (1978) Phytochemistry 17, 1343.
- Josephy, E. and Radt, F. (eds.). (1940) Elsevier's Encyclopaedia of Organic Chemistry, Vol. 14. Elsevier, New York
- 19. Kocoi, M. and St. Pyrek, J. (1973) J. Org. Chem. 38, 3688.
- Nagasampagi, B. A., Rowe, J. W., Simpson, R. and Goad, L. J. (1971) Phytochemistry 10, 1101.